

Multiple-Trait estimation of genetic parameters of yield traits of dairy cattle in Tunisia using an animal Model

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Abstract – A Markov Chain Monte Carlo Bayesian analysis was used on Tunisian dairy cattle data. Data included 14069 lactation records collected from 7139 animals over 16 freshening years from 1997 to 2013. The used model included fixed effects (herd-year, month of calving, and age-parity), a permanent effect, additive genetic effect, and a residual effect. (Co)-variance components were estimated by Bayesian methods. Gibbs Sampling was used to obtain conditional posterior distributions for additive, permanent environmental and residual variances and other genetic parameters. The variance of the residual effects represents the highest average between the three variance components, which is in the vicinity of 1493600 ± 25031 , 1833.4 ± 29.391 , and 1368.2 ± 22.074 , for milk, fat, and protein yields, respectively. Posterior means of heritability were 0.153 ± 0.018 , 0.11 ± 0.014 , and 0.13 ± 0.016 for the same traits, respectively. Posterior means of repeatability were in the range of 0.342 ± 0.011 , 0.252 ± 0.011 , and 0.31 ± 0.01 for milk, fat, and protein yields, respectively. The largest genetic correlation (0.94) was observed between milk and protein yields. These results should be useful to implement genetic evaluation for the Tunisian Holstein population. However, results also indicate that there should be additional focus on data recording quality.

Keywords: Markov Chain Monte Carlo, Bayesian Method, Gibbs Sampling, Posterior means, Genetic parameters

1. Introduction

Genetic selection in livestock populations and especially in dairy cattle programs is traditionally based on phenotypic records of the individual and its relatives (Meuwissen et al., 2001). Henderson's method 3 (Henderson, 1953) for estimating variance components was widely used until the late 1970's. With rapid advances in software technology, likelihood based methods gained favor in animal breeding. BLUP can be derived only if the dispersion parameters (variance and covariance components) are known (Sorensen and Gianola, 2002). In recent years, Bayesian methods have been developed for variance component estimation in animal breeding (Gianola and Fernando, 1986; Sorenson et al., 1994; Hallander et al., 2010). Bayesian analysis is gaining popularity because of its more comprehensive assumptions than those of classical approaches and its flexibility in resolving a wide range of biological problems (Waldmann, 2009; Hallander et al., 2010). In the Bayesian approach, the idea is to combine what is known about the statistical ensemble before the data are observed (prior probability distributions) with the information coming from the data, to obtain a posterior distribution from which inferences are made using the standard probability calculus techniques (Sorensen and Gianola, 2002; Robert, 2006). The size of the Holstein cow population has substantially increased over the recent years in Tunisia through the importation of pregnant heifer and semen (Ben Zaabza et al., 2016a). Hammami et al. (2008) reported that 60% of all inseminations of cows in Tunisia used Holstein semen. In Tunisia, breeding decisions are based on recorded yield or an intra-herd index for cows, and essentially on a milk yield index for AI bulls. However, breeding decisions do not take into account traits other than milk yield. There are four types of herds, the state herds, the cooperative herds, the groups of investors' herds, and the farmers' herds. These herds differ with feeding and management models and milk production levels (Rekik and Ben Gara., 2004; Ben Gara et al., 2006). Estimation of dispersion parameters in multipletrait is more challenging than in univariate mixed models because of the greater dimensions of multipletrait genetic evaluation systems. The estimation of genetic parameters is an important step in genetic



evaluation because they provide an indication of the capacity of a population to respond to selection, and thus, the potential of that population to evolve (Thomson and Hill, 2000a; Thomas et al., 2000b). The purpose of this study was to estimate genetic parameters of fat, protein, and milk yields in a multiple trait animal model using Bayesian analysis approach.

2. Materials and methods

2.1. Data

Data were provided by the National Centre for Genetic Improvement at Sidi Thabet, Tunis. Data consisted of 25765 completed lactation records collected from 1997 through 2013 on 9261 Holstein cows in 11 herds. All records included 305-d milk, fat, and protein yields. Each record included the international identification number, herd code, lactation number, calving date, milk yield, fat and protein percentages. The pedigree file included the sire, the dam, the date of birth, and the herd of origin for each animal. After editing for missing identification number, and unreasonable production levels for daily milk yield (<1.0 and > 50 kg), fat content (< 1.5% and > 5% and protein percentage (< 1% and >5%), 14069 records remained on 7193 cows sired by 326 bulls. Description of data structure is shown in table 1.

Table 1. Data structure							
Number		Mean 305-d yield(kg)	Cows in 1 st parity %				
Herd	Cows	Milk	Fat	Protein			
1	286	6010.54	210.477	188.71	31.88		
2	359	6348.88	203.73	192.092	33.15		
3	360	5641.61	184.93	176.63	37.66		
4	361	5668.95	189.103	172.103	41.49		
5	396	6491.14	220.23	194.30	29.01		
6	636	6929.83	229.24	216.21	36.77		
7	762	6583.44	226.293	213.306	43.81		
8	876	6851.03	266.52	236.89	35.40		
9	1016	6460.84	279.65	264.201	42.65		
10	1063	6018.22	214.53	194.16	39.6		
11	1078	6414.83	272.64	260.35	44.5		
Average	653.9	6310.846	227.031	209.904	37.81		

2.2. Statistical Model

The used mixed linear model for 3 traits y, z, and t in matrix notation was as follows:

$$\begin{bmatrix} y \\ z \\ t \end{bmatrix}_{=} \begin{bmatrix} X_{y} & 0 & 0 \\ 0 & X_{z} & 0 \\ 0 & 0 & X_{t} \end{bmatrix} \begin{bmatrix} \beta_{y} \\ \beta_{z} \\ \beta_{t} \end{bmatrix}_{+} \begin{bmatrix} W_{y} & 0 & 0 \\ 0 & W_{z} & 0 \\ 0 & 0 & W_{t} \end{bmatrix} \begin{bmatrix} P_{y} \\ P_{z} \\ P_{t} \end{bmatrix}_{+} \begin{bmatrix} Z_{y} & 0 & 0 \\ 0 & Z_{z} & 0 \\ 0 & 0 & Z_{t} \end{bmatrix} \begin{bmatrix} a_{y} \\ a_{z} \\ a_{t} \end{bmatrix}_{+} \begin{bmatrix} e_{y} \\ e_{z} \\ e_{t} \end{bmatrix}, (1)$$

where β_y , β_z and β_t are vectors of fixed effects affecting traits y, z, and t. P_y , P_z and P_t are vector of permanent effects order (s), and a_y , a_z and a_t are vectors of order q of additive genetic values for traits y, z, and t, and $e_y(e_z)$ is a vector of residual effects of order n(n) for the same traits y, z, and t. Fixed effects included herd-year, month of freshening, and age-parity. Matrices X, W, and Z, with subscripts y, z, and t are known incidence arrays relating location effects for each trait to data.

Put now $\beta = \begin{bmatrix} \beta_y & \beta_z & \beta_t \end{bmatrix}$, $P = \begin{bmatrix} P_y & P_z & P_t \end{bmatrix}$, $a = \begin{bmatrix} a_y & a_z & a_t \end{bmatrix}$ with appropriate partitions for matrices X, W, and $\begin{bmatrix} X_y & 0 & 0 \end{bmatrix} \begin{bmatrix} W_y & 0 & 0 \end{bmatrix} \begin{bmatrix} V_y & 0 & 0 \end{bmatrix}$

$$Z, \text{ such that } X = \begin{bmatrix} x_{y} & z_{t} \\ 0 & X_{z} & 0 \\ 0 & 0 & X_{t} \end{bmatrix}, W = \begin{bmatrix} x_{y} & z_{t} \\ 0 & W_{z} & 0 \\ 0 & 0 & W_{t} \end{bmatrix}, Z = \begin{bmatrix} z_{y} & z_{t} \\ 0 & Z_{z} & 0 \\ 0 & 0 & Z_{t} \end{bmatrix},$$

The conditional distribution of the complete data for each individual, given the parameters, is assumed to be multivariate normal and can be written as $Q|^{\beta, P, a, R_e} \sim MVN(X^{\beta+Za+WP, R}),$ (2)



where Q contains y, z, and t. We shall assume that records have been sorted by individual, so that Q is a sequence of y, z, and t for each individual. Hence, the sorting is such that the residual variance-

covariance matrix can be written $R = I_n \otimes R_e$ a block diagonal matrix with **n** sub-matrices of residual

$$\mathbf{R}_{e} = \begin{bmatrix} \sigma_{e,y}^{2} & \sigma_{e,yz} & \sigma_{e,yt} \\ \sigma_{e,zy} & \sigma_{e,z}^{2} & \sigma_{e,zt} \\ \sigma_{e,ty} & \sigma_{e,tz} & \sigma_{e,t}^{2} \end{bmatrix}$$

co-variances R_{e} , where R_{e} =

And, $\sigma_{e,y}^{e}$ is the residual variance for trait y, $\sigma_{e,z}^{e}$ is the residual variance for trait z, and $\sigma_{e,yz(t)}$ is the residual covariance.

2.3. Distribution a priori

Prior distributions are needed in Bayesian modeling. For the vector β , a proper uniform distribution is assigned, with density

 $P^{(\beta)\alpha}$ constant

The vector of additive genetic values is assumed to follow, a priori, the multivariate normal distribution. $a \downarrow G_{\theta}, A \sim MVN(0, G_0 \otimes A)$

where A is the additive genetic relationship matrix of order q×q, conditionally on an unknown genetic covariance matrix G_{θ} of order k×k and \otimes represents Kronecker product.

$$\boldsymbol{G}_{\boldsymbol{\theta}} = \begin{bmatrix} \boldsymbol{\sigma}_{a,y}^{2} & \boldsymbol{\sigma}_{a,yz} & \boldsymbol{\sigma}_{a,yt} \\ \boldsymbol{\sigma}_{a,zy} & \boldsymbol{\sigma}_{a,z}^{2} & \boldsymbol{\sigma}_{a,zt} \\ \boldsymbol{\sigma}_{a,ty} & \boldsymbol{\sigma}_{a,tz} & \boldsymbol{\sigma}_{a,t}^{2} \end{bmatrix}$$

 $\sigma_{a,y}$ is the variance between additive genetic effects affecting trait 1, and $\sigma_{a,yz(t)}$ is the additive covariance between traits. Similarly, the prior distribution of P is also multivariate normal.

$$P \mid \boldsymbol{R}_{p} \sim MVN^{(0, R_{p} \otimes I_{S})}, \text{ where } R_{p} = \begin{bmatrix} \sigma_{p,y}^{2} & \sigma_{p,yz} & \sigma_{p,yt} \\ \sigma_{p,zy} & \sigma_{p,z}^{2} & \sigma_{p,zt} \\ \sigma_{p,ty} & \sigma_{p,tz} & \sigma_{p,t}^{2} \end{bmatrix}$$

The prior distributions of covariance matrices G_0, R_p , and R_e are each k-dimensional inverse Wishart. Three-dimensional scaled inverted Wishart distributions were assigned as prior processes for each of the R_e, R_p, G_0 co-variance matrices, with the respective densities being.

$$\begin{array}{l} P(R_{e} | v_{e}, V_{e}) \alpha & |R_{e}|^{-\frac{1}{2}(V_{e}^{+k+1})} & -\frac{1}{2}tr(R_{e}^{-1}V_{e}^{-1}) \\ P(R_{e} | v_{p}, V_{p}) \alpha & |R_{p}|^{-\frac{1}{2}(V_{p}^{+k+1})} & -\frac{1}{2}tr(R_{p}^{-1}V_{p}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) \alpha & |G_{a}|^{-\frac{1}{2}(V_{a}^{+k+1})} & exp(-\frac{1}{2}tr(G_{0}^{-1}V_{a}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) & |G_{a}|^{-\frac{1}{2}(V_{a}^{+k+1})} & exp(-\frac{1}{2}tr(G_{0}^{-1}V_{a}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) & |G_{a}|^{-\frac{1}{2}(V_{a}^{+k+1})} & exp(-\frac{1}{2}tr(G_{0}^{-1}V_{a}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) & |G_{a}|^{-\frac{1}{2}(V_{a}^{+k+1})} & exp(-\frac{1}{2}tr(G_{0}^{-1}V_{a}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) & |G_{a}|^{-\frac{1}{2}(V_{a}^{+k+1})} & exp(-\frac{1}{2}tr(G_{0}^{-1}V_{a}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) & |G_{a}|^{-\frac{1}{2}(V_{a}^{+k+1})} & exp(-\frac{1}{2}tr(G_{0}^{-1}V_{a}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) & |G_{a}|^{-\frac{1}{2}(V_{a}^{+k+1})} & exp(-\frac{1}{2}tr(G_{0}^{-1}V_{a}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) & |G_{a}|^{-\frac{1}{2}(V_{a}^{+k+1})} & exp(-\frac{1}{2}tr(G_{0}^{-1}V_{a}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) & |G_{a}|^{-\frac{1}{2}(V_{a}^{+k+1})} & exp(-\frac{1}{2}tr(G_{0}^{-1}V_{a}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) & |G_{a}|^{-\frac{1}{2}(V_{a}^{+k+1})} & exp(-\frac{1}{2}tr(G_{0}^{-1}V_{a}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) & |G_{a}|^{-\frac{1}{2}(V_{a}^{+k+1})} & exp(-\frac{1}{2}tr(G_{0}^{-1}V_{a}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) & |G_{a}|^{-\frac{1}{2}(V_{a}^{+k+1})} & exp(-\frac{1}{2}tr(G_{0}^{-1}V_{a}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) & |G_{a}|^{-\frac{1}{2}(V_{a}^{+k+1})} & exp(-\frac{1}{2}tr(G_{0}^{-1}V_{a}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) & |G_{a}|^{-\frac{1}{2}(V_{a}^{+k+1})} & exp(-\frac{1}{2}tr(G_{0}^{-1}V_{a}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) & |G_{a}|^{-\frac{1}{2}(V_{a}^{+k+1})} & exp(-\frac{1}{2}tr(G_{0}^{-1}V_{a}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) & |G_{a}|^{-\frac{1}{2}(V_{a}^{+k+1})} & exp(-\frac{1}{2}tr(G_{0}^{-1}V_{a}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) & |G_{a}|^{-\frac{1}{2}(V_{a}^{+k+1})} & exp(-\frac{1}{2}tr(G_{0}^{-1}V_{a}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) & |G_{a}|^{-\frac{1}{2}(V_{a}^{+k+1})} & exp(-\frac{1}{2}tr(G_{0}^{-1}V_{a}^{-1}) \\ P(G_{a} | v_{a}, V_{a}) & |G_{a}|^{-\frac{1}$$

where k = 3. In these expressions, ^Vi and Vi (i=e, p, a) are hyperparameters of the distributions, which are assumed known. The marginal distribution for each parameter is found via integration of multivariate density functions using a Gibbs sampling technique as developed by Sorensen and Gianola (2002).

2.4. Gibbs Sampling

In a standard multiple-trait analysis, all full conditional distributions needed for Bayesian implementation of a MCMC algorithm, such as the Gibbs sampler (García-Cortés and Sorensen, 1996; Waldmann et al., 2009). Variance components were estimated with a Bayesian approach via the Gibbs sampling algorithm as implemented by Misztal et al. (2002). Posterior means of variance components,



heritability, and correlation estimates were obtained using 50,000 samples. After a burn-in of 5000 samples, and then one out of 10 iterations was kept for subsequent analysis Thus, a total of 4500 samples were saved. Convergence of Gibbs chains was monitored by inspection of plots related to selected parameters.

3. Results and Discussion

Summary statistics of distributions, the mean, median, and variance and correlations among production traits are reported in table 2-5. The estimated marginal posterior densities of heritability estimates are shown in figure1. Typically, posterior densities of genetic parameters were not symmetric, and associated with a reasonable difference between mean, mode, and median. Genetic and permanent environment correlations between milk, fat, and protein yields were close to 1.00. Genetic correlations between milk and fat, milk and protein, and fat and protein yields were 0.81, 0.94, and 0.86, respectively. Permanent environment correlations between yield traits were larger than the corresponding genetic correlations, except for protein with fat yields. Heritability estimates from this analysis were 0.15, 0.11, and 0.13 for milk, fat, and protein yields, respectively, with standard errors of approximately 0.01. Posterior means of repeatability (Tables 3 and 4) were similar for milk and protein yields (0.3), but repeatability for fat yield tended to be slightly lower than those for milk yield (0.25). The additive variance of the milk yield ranged from 204000 kg² to 492000 kg². However, the additive variance of the fat yield ranged from 228 kg² to 300 kg². Permanent environmental variances were larger than additive genetic variance for milk, fat, and protein yields, indicating that environmental effects had a higher impact on the variation of milk production. For all traits, estimated permanent environmental variances were around 1.3 times higher than the additive genetic variances. The residual variances were high in comparison with genetic additive and permanent environmental variance for 305-d milk, fat, and protein vields. Estimated genetic correlations among 305-d milk, fat, and protein yields were comparable with those estimated by Hammami et al. (2008b), and Muir et al. (2007). However, the genetic correlation between milk and fat yields was larger than those obtained by Carabaño et al. (1989) on United States data. Estimates of the genetic correlations between milk and protein yields were also somewhat greater than the estimate (0.88) reported by Van Vleck and Dong (1998). Estimates of heritability for milk ranged from 0.13 to 0.21, these results are similar to those observed from study used a 305-d model as applied in this study (Ben Gara et al., 2006; Ben Zaabza et al., 2016a). Values obtained in this study for heritability of fat-yields are similar in magnitude and trend to those observed by Carabaño et al. (1989). However, heritability of Milk, fat, and protein yields were smaller than those found in the literature (Ahlborn and Dempfle, 1992; De Ross et al., 2004; Muir et al., 2007). Considerable variation exists between countries for genetic parameters estimates related to herd management systems (De Veer and Van Vleck, 1987; Zwald et al., 2001; Ojango et al., 2002). Smaller genetic variances and heritability for Tunisian dairy cattle population than for the other Holstein populations can be explained by the lowest production levels and stressful climatic conditions (Veerkamp et al., 1998; Ojango et al., 2002; Muasya et al., 2014). In fat, In Tunisia, the climate varies from arid in the South to humid in the North, and characterized by hot summers coupled with high humidity (Ben Zaabza et al., 2016b). In Tunisia, the feeding system is unbalanced and rations are based mainly on concentrates. The forage is characterized by poor quality, and high rate in indigestible cellulosic constituents that could be possible causes of the lower milk yield. All these factors could lead to decreases in production performances and increase in the incidence of health troubles such as acidosis at the herd level. Increased heat stress can severely depress milk production (Huquet et al., 2012; Hammami et al., 2015). Posterior mean estimates of additive genetic variance of 305-d milk yields is 2 times lower than that obtained by Misztal et al. (1992) and Carabaňo et al.(1989) analyzing 305 d milk yield on US Holsteins. However, additive genetic estimates in our study were larger than those estimated by Ojango et al. (2002) in the Kenyan data (349350 kg² vs. 221797). Ojango et al. (2002) estimated genetic parameters for 305-d milk yield using bivariate animal model analysis in the Kenya and UK. They reported a difference in genetic variance between two countries (221797 kg² in Kenya vs. 582537 in UK). Posterior means of repeatability (Tables 3 and 4) were similar for milk and protein yields (0.3), but repeatability for fat yield tended to be slightly lower than those for milk yield (0.25). For milk yield, a similar pattern was observed by Ben Gara et al. (2006) in the same population using earlier records. Moreover, the additive variance of the fat yield ranged from 228 kg² to 300 kg². Estimates obtained by Campos et al. (1994) were 366.634 kg² for milk yield and 516 kg² for fat yield of Holstein cattle in Florida using derivative-free REML with the animal model. The difference between estimates in this study and those of literature may be partially



due to inaccurate pedigree information on imported semen of some sires (Hammami et al., 2008b), and the lowest average number of daughters per bull. Estimates of residual variance for milk, fat, and protein yields were 1493600, 1833.4, and 1368.2 kg², respectively. These estimates were larger than those estimated by Sun et al. (2009) using a multiple-trait model including female fertility and milk production traits. They reported residual variance estimates of 955216, 1504.3, and 841.6 kg² for milk, fat, and protein yield, respectively. Uncontrolled environmental factors might be a possible explanation for this observed trend. However, observed values for residual variance were lower than those obtained in the previous study on a sample of the Iranian Holstein dairy cattle using Bayesian procedure (Alijani et al., 2012). Component of the residual variance could include the whole of the factors not yet taken into account, which they are of genetic origin (nonadditive genetic effects) or related to the environment (factor not identified such as the reproductive state of the animal). THomas and Hill (2000) reported that missing pedigree information and incorrectly assigned relationships can cause larger bias in estimates of genetic parameters and variance components.



Figure 1. Estimated marginal posterior densities of heritability (h²), and repeatability (r) of milk, fat, and protein yields.



Table 2. Genetic (above diagonal) and permanent environmental (below diagonal) correlations (SD in brackets) for 305-d milk, fat and protein yields.

Trait Milk yield 305-d	Milk yield 305-d	Fat yield 305-d 0.81(0.01)	Protein yield 305-d 0.94(0.01)
Fat yield 305-d	0.82(0.01)		0.86(0.01)
Protein yield 305-d	0.95(0.02)	0.85(0.03)	

Table 3. Summary of the marginal distributions of additive genetic, permanent environment (PE) and residual (co)variance components, heritability, repeatability associated to 305-d milk yields

Table 4. Summary of the marginal distributions of additive genetic, permanent environment (PE) and residual (co)variance components, heritability, repeatability associated to 305-d fat yields.

Parameter Additive genetic variance	Mean 268.79	SD 37.00	Min 204.90	Max 342.50	Median 267.20	Mode 241.89
PE variance	349.91	36.32	281.30	422.40	350.30	346.54
Residual variance	1833.4	29.39	1777.00	1891.0	1833.00	1836.5
Heritability	0.11	0.014	0.086	0.184	0.109	0.109
Repeatability	0.252	0.011	0.267	0.349	0.252	0.235

Table 5. Summary of the marginal distributions of additive genetic, permanent environment (PE) and residual (co)variance components, heritability, repeatability associated to 305-d protein yields.

Parameter	Mean	SD	Min	Max	Median	Mode
Additive genetic variance	262.67	34.40	202.90	332.9	260.90	241.26
PE variance	352.13	33.046	289.00	416.60	353.00	356.14
Residual variance	1368.20	22.074	1328.00	1414.00	1368.00	1366.40
Heritability	0.132	0.016	0.086	0.184	0.131	0.109
Repeatability	0.31	0.01	0.267	0.342	0.31	0.319

4. Conclusion

Genetic parameters of milk, fat, and protein yields were estimated for Tunisian Holsteins using a Multivariate Bayesian analysis. Heritability and repeatability were moderate but similar to previous literature estimates from studies that used a comparable model in the same population, indicating the possibility of a satisfactory response to selection for these production traits in Tunisian Holsteins. Genetic parameters estimates in this study might be used in the official genetic evaluation for production traits for Tunisian Holsteins.

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