

Effects of some Essential Oils and Fungal Isolates on Mycelial Growth of *Fusarium oxysporum*: a Soilborne Pathogen Isolated from Tunisian Geothermal Greenhouses

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Abstract – Four essential oils (EOs) and four antagonist fungi, isolated from date palm compost, were tested against *Fusarium oxysporum*, the causal agent of tomato wilt and fruit rot in Tunisian geothermal greenhouses. EOs have shown anti-*F. oxysporum* activity ranged between 36.94 and 100%. *Lavandula angustifolia* EO exhibited the highest antifungal activity at 5µl/ml marked by complete mycelial growth inhibition of the pathogen. With the exception of *Cymbopogon citrates* EO which showed, statistically, the same activity even at the low concentration, a significant difference was observed between concentrations for the other EOs. Inhibition of *F. oxysporum* mycelial growth was also obtained in the presence of *Aspergillus Candidus*, *A. Terreus*, *Talaromyces assientensis* and *Penicillium chrysogenum* which showed the best efficacy (50.37%). Antibiosis and/ or parasitism are the two main mechanisms employed by the tested antagonists in their antifungal activity.

Keywords: Antagonists, Antifungal activity, Essential oils, *Fusarium oxysporum*.

1. Introduction

In order to reduce losses caused by soilborne pathogens and achieve high crop yields, pesticides are widely used in agricultural. Excessive application and irrational use of those synthetic products have significant drawbacks and have been proved to be harmful to human health and environment (Paster and Bullerman, 1988). Thus increase interest has been focused on the development of healthy and safer alternatives to replace chemical pesticides. One such alternative is the use of natural antifungal agents such as plant EOs and potential antagonists to control plant pathogens (Costa et al. 2000).

EOs are characterized by their easily extractability, eco-friendly, biodegradable and for their effectiveness against wide spectra of pests (Isman, 2000; Lucia et al. 2012). Several microorganisms isolates with potential antagonist effect against soilborne pathogenic fungi have been also described (Howell, 2003; Faltin et al. 2004).

Biological control agents have proven to have specific advantages compared to synthetic fungicides. In addition of their safety environmental effects, a reduced probability of resistance pathogens development was also noted (Cook, 1988).

In this project, we studied the biofungicide effect of some EOs and fungi isolates against *F. oxysporum*, a pathogen isolated from Tunisian geothermal greenhouses.

2. Materials et Methodes

2.1. Pathogen

The pathogen used in this study was isolated from diseased tomato plants grown under geothermal greenhouse conditions and showing typical wilting symptoms of *Fusarium oxysporum*.

Fungal pathogen was isolated by planting infected tissues on Potato Dextrose Agar (PDA) medium supplemented with streptomycin-sulphate at 250 mg/ml. The pathogen was purified using the single spore culture and the culture plates were incubated at 28°C for 5 days.



2.2. Test microorganisms and growth medium

Four antagonist fungal species, isolated from compost, were used as biocontrol agents, namely: *Aspergillus terreus*, *A. candidus*, *Penicillium chrysogenum* and *Talaromyces assientensis* (El Khaldi et al. 2016). Fungal strains were cultured and maintained on Potato Dextrose Agar (PDA)

2.3. Test essential oils

Four EOs: *Lavandula angustifolia* (*L. angustifolia*), *Origanum majorana* (*O. majorana*), *Santolina chamaecyparissus* (*S. chamaecyparissus*) and *Cymbopogon citratus* (*C. citratus*) were kindly obtained from the Regional Center for Horticulture and Organic Agriculture (Laboratory of Entomology) Chott–Mariem, Sousse, Tunisia.

2.4. Antifungal assay

Biofungicide effects of EOs were assessed on PDA medium at concentrations of 0.2, 0.5, 2.5 and 5 µl/ml. These concentrations were obtained by the addition of 4, 10, 50 and 100 µl of EO in 20 ml of the lukewarm PDA. After stirring, the medium was poured into 9 cm Petri dishes. 6 mm disc of 5 old culture of the pathogen was placed in the middle of each plate and then incubated at 28°C for 7 days.

Inoculated PDA without EOs served as a control. Data were expressed as percentage inhibition of mycelial growth relative to the control. Antifungal index was determined by the formula:

$I\% = 100 \times (r_1 - r_2)/r_1$ as described by Hmouni et al. (1996) where r_1 and r_2 = mean radial growth of pathogen in absence or presence of EO respectively.

Antagonistic activity of fungal isolates against pathogen was tested according dual culture method. 6 mm discs of pathogen and each fungal isolate removed from the colony margin of actively growing cultures on PDA were placed equidistantly in the same Petri dish. Plates were then incubated in the dark at 28 °C for 7 days. The percentage inhibition of radial growth (I) of the pathogen was determined as: $I\% = 100 \times (r_1 - r_2)/r_1$ as described by Hmouni et al. (1996) where r_1 and r_2 = mean radial growth of pathogen in absence or presence of antagonist respectively.

2.5. Statistical analysis of data

Three plates were used per experiment for each treatment of biocontrol agent or EOs, and the whole experiment was repeated twice. Statistical analysis were performed with the STATISTICA software version5 and comparisons of group means were accomplished with the Newman–Keuls test at $P \leq 0.05$. Linear regression analyses were performed to establish any correlations among different concentrations of EOs and their antifungal activity.

3. Results and Discussion

3.1. Antifungal activity of essential oil against *Fusarium oxysporum*

The antifungal activity of EOs on *F. oxysporum* was assessed by direct contact method. The obtained results are reported in Fig 1.

Mycelial growth inhibition of *F. oxysporum* was observed for the four EOs and at all tested concentrations. The range of mycelia growth inhibition was between 36.94 and 100%.

Overall, as the concentration of EO increased, the antifungal activity increased. The inhibitory effect of *O. majorana* showed dose-dependent activity. Results demonstrated significant differences ($P \leq 0$) between the tested concentrations with lowest activity (36.94%) recorded at 0.2 µl/ml. For *L. angustifolia* and *S. chamaecyparissus* EOs, no significant difference was observed between 0.2 and 0.5 µl/ml as well as for 2.5 and 5 µl/ml. In case of *C. citrates* EO treatment, dose-response effect on the growth of *F. oxysporum* was not detected. In fact no significant differences at $P \leq 0$ were observed for the different concentrations and the mycelia growth inhibition was ranged between 79.80 to 92.65%.

The relationship between dose response effect of the different EOs and the growth of *F. oxysporum* was assessed by correlation analysis (Fig 2). There was a significant positive correlation ($P \leq 0.05$) between the tested EO concentrations and the mean radial growth inhibition of *F. oxysporum*: *L. angustifolia* ($R^2 = 0.84$), *O. majorana* ($R^2 = 0.96$), *S. chamaecyparissus* ($R^2 = 0.85$) and *C. citratus* ($R^2 = 0.68$).

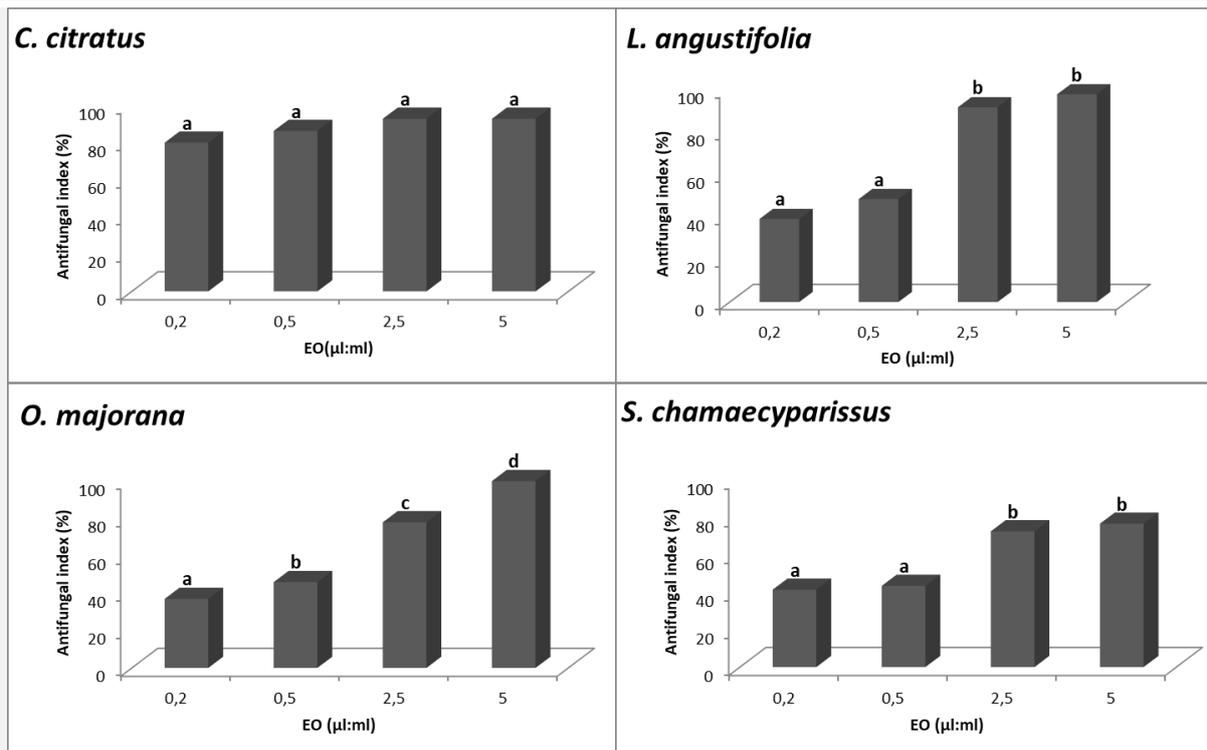


Figure 1. Antifungal index (%) of *L. angustifolia*, *O. majorana*, *S. chamaecyparissus* and *C. citratus* essential oils against *F. oxysporum* at different concentration after 7 days of incubation at 28°C. Means followed by same letter are not significantly different from each other at $P \leq 0.05$ based on Newman-Keuls test.

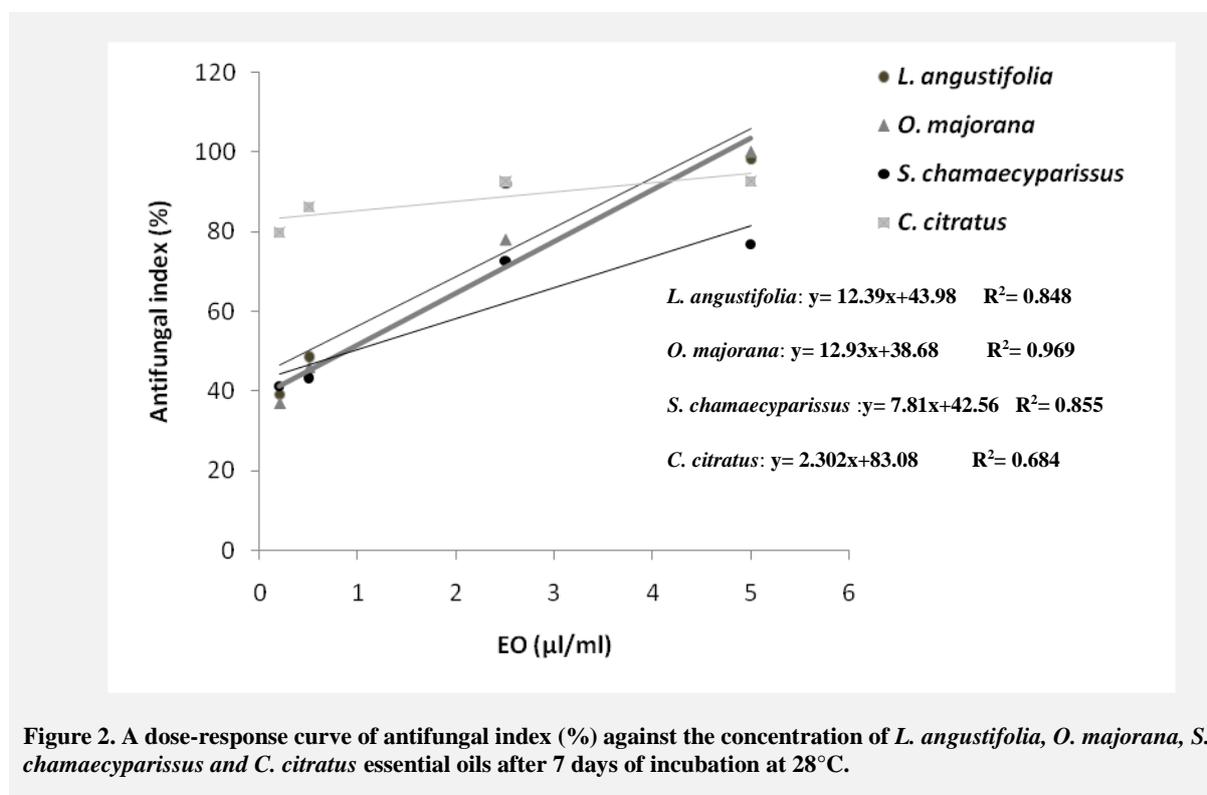


Figure 2. A dose-response curve of antifungal index (%) against the concentration of *L. angustifolia*, *O. majorana*, *S. chamaecyparissus* and *C. citratus* essential oils after 7 days of incubation at 28°C.

Comparison between tested EOs was further confirmed by comparing their effectiveness at the same concentration on the growth of *F. oxysporum* (Fig 3). Results showed that for the minimum concentration as well as for 0.5µl/ml concentration, with the exception of *C. citratus*, all the rest of EOs

showed statically ($P \leq 0.05$) the same antifungal effect. For 2.5 $\mu\text{l/ml}$ concentration, *L. angustifolia* and *C. citratus* have the highest effect (91.84 and 92.65%) and they are not significantly different at $P \leq 0.05$. At 5 $\mu\text{l/ml}$, *O. majorana* and *L. angustifolia* EOs, which have significantly ($P \leq 0.05$) the same antifungal activity against the target, revealed the higher values of inhibition (100 and 97.96%, respectively) (Fig 4). 92.65% of radial growth inhibition was recorded with *C. citratus* EO and 76.73% with *S. chamaecyparissus* EO.

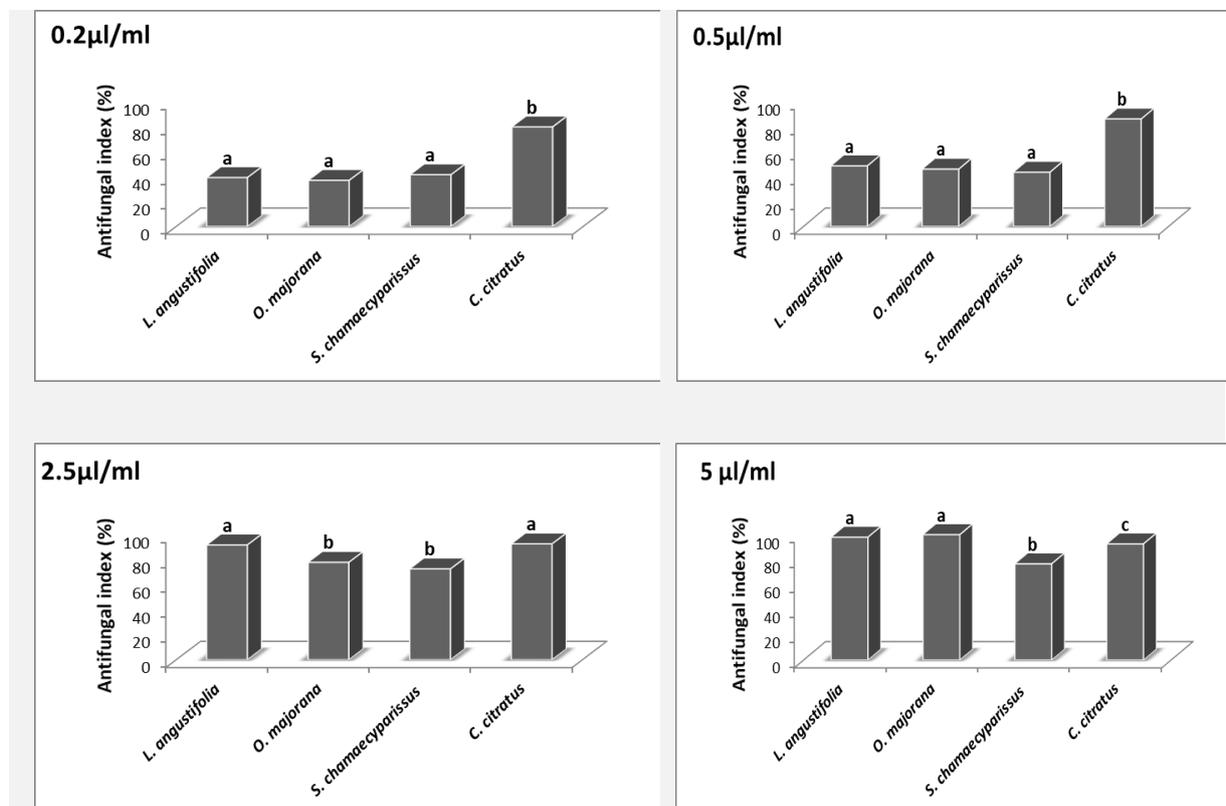


Figure 3:Antifungal effectiveness between *L. angustifolia*, *O. majorana*, *S. chamaecyparissus* and *C. citratus* essential oils against *F. oxysporum* at different concentration after 7 days of incubation at 28°C. Means followed by same letter are not significantly different from each other at $P \leq 0.05$ based on Newman-Keuls test.

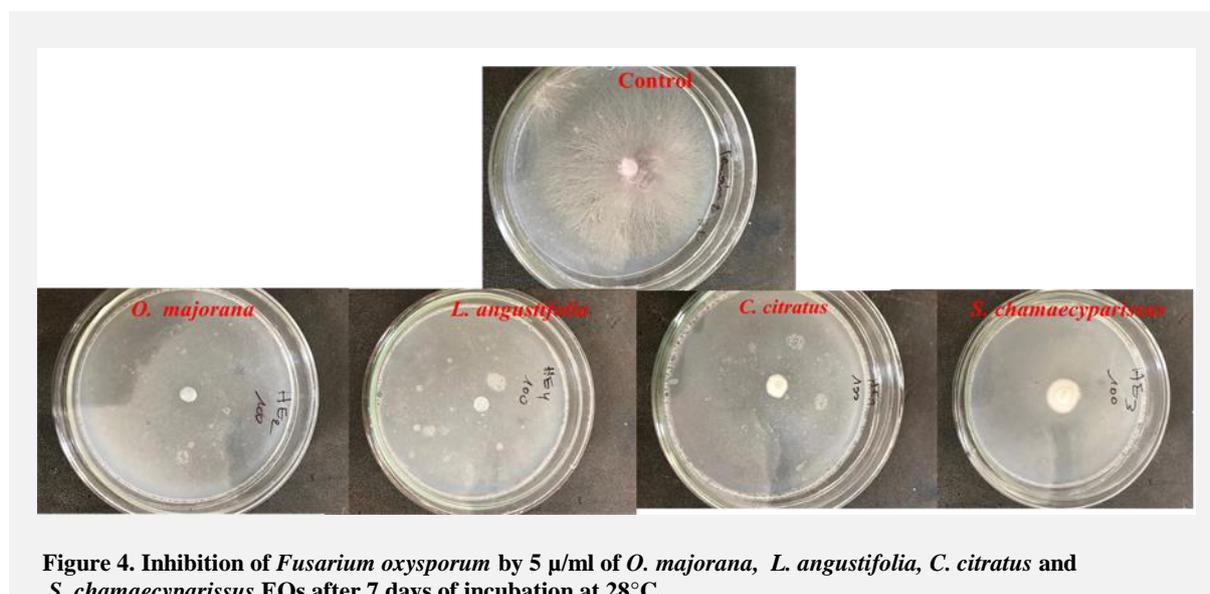


Figure 4. Inhibition of *Fusarium oxysporum* by 5 $\mu\text{l/ml}$ of *O. majorana*, *L. angustifolia*, *C. citratus* and *S. chamaecyparissus* EOs after 7 days of incubation at 28°C.

Four compost isolates were tested against *F. oxysporum* in dual culture plates (Fig 5). After 7 days at 28 °C, results of the antagonism tests showed that mycelia growth was inhibited an average of 18.86 % for *T. assientensis*, 47.54% for *A. Terreus*, 48.49% for *A. Candidus* and 50.37% for *P. chrysogenum*, which was the most effective biocontrol agent. Two different mode of inhibition to pathogen was observed: antibiosis and parasitism. No physical colony contact was observed between *T. assientensis* and *F. oxysporum* and an inhibition zone was observed, which indicates the presence of fungistatic metabolites secreted by the antagonist. For the remains fungi, they are able to overgrow the mycelium of *F. oxysporum*.

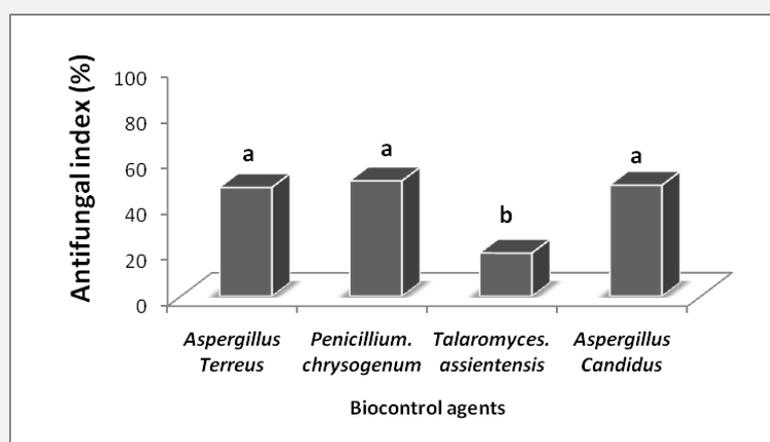


Figure 5. Antifungal index (%) of *Aspergillus terreus*, *Aspergillus candidus*, *Penicillium chrysogenum* and *Talaromyces assientensis* against *Fusarium oxysporum* after 7 days of incubation at 28°C.
Means followed by same letter are not significantly different from each other at $P=0.05$ based on Newman-Keuls test

In recent years, there has been a growing interest in researching and developing of eco-friendly and sustainable alternatives to control phytopathogens in agriculture using natural antifungal agents (Costa et al. 2000). In this context, the aim of the current study was to evaluate the antifungal activity of selected EOs and biocontrol agents against *F. oxysporum*.

EOs extracted from *L. angustifolia*, *O. majorana*, *S. chamaecyparissus* and *C. Citrates* showed anti-*F. oxysporum* activity. All tested EOs concentrations are able to inhibit mycelia growth of the fungal. Previous studies reported that *C. citratus* essential oil has the potential for the control of *Fusarium* spp. in vitro. *C. citratus* EO decreased seven different species of *Fusarium*, growth in PDA even at the lowest concentration (Gawai, 2015) being in accordance with the present study. Gupta et al. (2011) and Manganyi et al. (2015) has obtained a completely inhibition growth of the isolate *F. oxysporum* by EO of *C. citrates* at some tested concentration. Strong antifungal effect against *F. verticillioides* was also observed at 500 and 1000 ppm *C. citratus* EO (Mishra and Dubey, 1994). This EO was found to be effective also for controlling other various range of plant pathogenic fungi such as *A. niger* (Baratta et al. 1998) *A. flavous* (Paranagama et al. 2003) and *Cladosporium* sp (Singatwadia and Katewa, 2001). Tzortzakis and Economakis (2007) revealed that fungal spore production of *Colletotrichum coccodes*, *Botrytis cinerea*, *Cladosporium herbarum*, and *Rhizopus stolonifer* was inhibited by *C. citrates* EO.

Although *C. citratus* was the most effective EO at all concentrations, *L. angustifolia* and *O. majorana* showed the strongest antifungal activity at 5 μ l/l.

The effectiveness of *Lavandula* sp and *Origanum* sp EO has been proven in several studies against a variety of fungi. Antifungal activity of *Origanum vulgare* and *Lavandula angustifolia* essential oils was demonstrated against *Aspergillus niger*, *A. ochraceus*, *Penicillium* sp. and *Trichoderma viride* (Stupar et al. 2014).

Verticillium dahliae and *Penicillium aurantiogriseum* have shown a high sensitivity to the *O. majorana* EO and the mycelial growth was strongly inhibited for both species of fungi even at small concentrations (Rus et al. 2015). Angioni et al. (2006) reported the same tendency in an oil extract from *Lavandula stoechas* L. ssp. *Stoechas* for the inactivation of *Rhizoctonia solani* and *F.oxysporum*.

Despite its less activity compared to the others tested EOs, *S. chamaecyparissus* exhibited appreciable anti- *F.oxysporum* effect in our study. Assessed against *F. solani*, Khubeiz and Mansour (2016) showed

the strong mycelial growth reduction with minimum inhibitory concentration (MIC) of about 0.8 mg/ml. While Nouasri et al. (2015) did not find any activity of this EO against *Aspergillus flavus* and moderate inhibition zone for *Fusarium* and *Penicillium* with higher MIC.

The antifungal activity of the fourth Tunisian EOs has been shown in our study. Variation in effectiveness can be attributed to difference in chemical composition between EOs (Soylu et al. 2010), structural configuration of the constituent components of the volatile oils and their functional groups (Kim and Park, 2012). Many scientific investigations have been conducted to identify the main components of EOs with antifungal activity. It was proved that the composition of EO varies because of different species and chemotypes (Knobloch et al. 1989; Tantaoui-Elaraki et al. 1993), and there is a hard link between the chemical structures of the most abundant compounds in EOs and the antifungal effect (Felšöciová et al. 2015).

Four compost indigenous fungi isolates, were also studied for their suitability as biocontrol agents against *F. oxysporum*. In vitro antagonism tests have shown the effectiveness of isolates in reduction of pathogen mycelia growth. Other studies performed with this species reported a potential in the control of *R. solani* with total inhibition (100%) of mycelial growth noted for *A. candidus* (El Khaldi et al. 2016). Mejdoub-Trabelsi et al. (2016) tested ten nonpathogenic *Aspergillus* spp. and *Penicillium* spp. isolates against *Fusarium sambucinum*, *F. oxysporum* and *F. graminearum* and reported a growth inhibition rate varied from 32 to 59%.

4. Conclusion

The data reported in this study show that EOs and selected fungal strains have antagonist effect against *F. oxysporum*. Superior effectiveness of EOs, compared to the isolates, was noted. These results might provide promising eco-friendly alternatives to the toxic fungicides for plant disease management.

However, further investigations are required for: identifying the most active antifungal compounds and secondary metabolites involved in this antagonistic activity, determining the mechanisms of action and quantifying their effect on disease incidence and severity.

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