

# The effect of acetic acid treatment on germination parameters of carrot seeds infected with “*Candidatus Liberibacter solanacearum*”

S. BEN OTHMEN<sup>1</sup>, M. ELIMEM<sup>2\*</sup>, H. ABIR<sup>1</sup>, K. ABBES, B. CHERMITI<sup>1</sup>

<sup>1</sup> High Agronomic Institute of Chott-Mariem, University of Sousse, 4042 Chott-Mariem, Tunisia

<sup>2</sup> Higher School of Agriculture, Mograne, Carthage University, 1121 Zaghuan, Tunisia

\*Corresponding author: mohammed.elimem123@gmail.com

**Abstract** – The purpose of the research was to evaluate the antibacterial activity of acetic by comparing certain germination parameters of two carrot seed lots. The first seed lot is infected with the fastidious plant pathogenic bacterium ‘*Candidatus Liberibacter solanacearum*’ recently detected in Tunisia while the second one was safe. Contaminated seeds were treated with acetic acid solutions at different concentrations of 5%, 10% and 20%. Seeds were tested on moist filter paper, in seed trays under controlled conditions. Results showed a clear significant difference between tested germination parameters of both lots of seeds, explaining thus the negative effect of this fastidious bacterium on germination parameters. Acetic acid treatment has a significant positive effect on germination parameters of contaminated carrot seeds. However, at the highest concentration, these parameters were negatively affected. These results may help carrot producers to develop a control strategy able to minimize the spread of this bacterium.

**Keywords:** organic acid, alternative treatments, seed borne pathogen, germination parameters

## 1. Introduction

Plant pathogens are considered as one of the most serious problems in cultivation since they can reduce the quantity and the quality of harvested seeds. In fact, some plant pathogens could be preserved in seed lots; in this way, seeds can inadvertently provide an efficient means of plant pathogen dissemination. The bacterium ‘*Candidatus Liberibacter solanacearum*’ (Liefting et al. 2009), also known as ‘*Ca. Liberibacter psyllaurosus*’ and referred to as CaLso (Hansen et al. 2008), is associated with zebra chip, which is one of the most economically important bacterial diseases of potato (*Solanum tuberosum* L.). More recently CaLso was also shown to cause vegetative disorders in Apiaceous crops in different areas worldwide (Haapalainen 2014). According to (Pitman et al. 2011) CaLso is transmitted from potato mother tubers to growing plants as well as to progeny tubers. For carrot crops, this bacterium was also considered as a carrot seed borne pathogen according to (Bertolini et al. 2014); being a seed borne disease enhance the risk of its introduction into new areas in which contaminated carrot seeds are commercialized.

The most effective means of controlling seed borne diseases is essentially the exclusion or reduction of the inoculum during seed production. Unfortunately, infection and contamination cannot always be avoided. Various seed treatments are used to reduce the amount of inoculum transmitted by the seeds. Seed treatments are strongly favoured over field sprays, as relatively small amounts of active compounds are required and applications can be made in restricted areas in order to reduce health risks. Among the used treatments there may be mentioned some physical treatments such as: hot water, electron beam and hot humid air that are commercially used for seeds disinfection. These treatments are able to reduce the inoculum load, but they are often not able to completely eliminate the inoculum without affecting the vitality of the seeds (Forsberg et al. 2002). Natural antimicrobial compounds could be also used as another alternative for seed disinfection, thus they can be combined with physical treatment in order to enhance their efficacy.

In fact, various organic acids exhibit antimicrobial activity (Bloukas et al. 1997). Some of them, such as acetic acid and lactic acid have been used for controlling seed borne diseases as they present low ecotoxicological profiles. According to El-Naimi et al. (2000), Saidi et al. (2001) and Borgen et al. (2004) acetic acid and lactic acid reduced infection of common bunt in wheat from 64% to 96%.

Since seed health has a particular importance and seeds are considered as a potential vehicle for transmitting beneficial or deleterious bacteria, this paper focuses on the study of the effect of CaLso on



some germination parameters for carrot seeds, especially the germination test as it is considered as the most important quality in evaluating the planting value of seed lot. The effect of acetic acid at different concentration was also studied in order to determine the suitable concentration for reducing the effect of CaLso and preserving seed viability.

## 2. Material et Methods

### 2.1. Carrot seeds

Seed samples of the cultivar “Arbi Zaafrana” were obtained from carrot seeds production area (Zaafrana, Kairouan, Tunisia), during two consecutive years 2015 and 2016. In 2015, three lots of seeds (SE1, SE2 and SE3) were analyzed for CaLsol presence and they tested negative (Ben Othmen et al. 2018). In 2016, seed samples (SE4, SE5 and SE6) were proved to be highly contaminated with a viability of CaLso cells ranging from 36% to 65% (Ben Othmen et al. 2018).

### 2.2. Seed treatment

The effect of seed treatment with acetic acid on CaLso on carrot seeds was assessed. Contaminated seed samples of 2016 were treated with acetic acid solutions at concentrations of 5%, 10% and 20%. For each treatment seeds were soaked in each solution for ten minutes then, they were placed in a sieve and excess compound was removed with sterilized distilled water. Thus, they were placed in plates to dry overnight at room temperature.

### 2.3. Seed germination test

Viable seeds with uniform size were selected and used for germination test. Three replicates of 50 seeds from each treatment (150 seeds) were placed in labeled Petri dishes containing two layers of moistened blotters and incubated at 28°C, in darkness. The total number of germinated seeds (Gmax) was determined on the basis of the number of seeds with visible radical ( $\geq 2$  mm) counted daily for 10 days. Seeds were considered germinated when there was a visible coleoptiles protrusion through the testa. The following germination parameters were recorded:

**Germination percentage (Gp)** = (# seeds sprouted/ #total seeds sprouted) x 100

### Mean Germination Time

$$\text{MGT (day)} = \sum f \times x / \sum f \text{ (Orchard 1977)}$$

(Where f=Seeds germinated on day x)

### Coefficient of Velocity of Germination

$$\text{CVG} = \frac{G_1 + G_2 + \dots + G_n}{(1 \times G_1 + 2 \times G_2 + \dots + n \times G_n)}$$

### The germination index

(GI) was calculated as described in the Association of Official Seed Analysts (AOSA, 1983) by following formula:

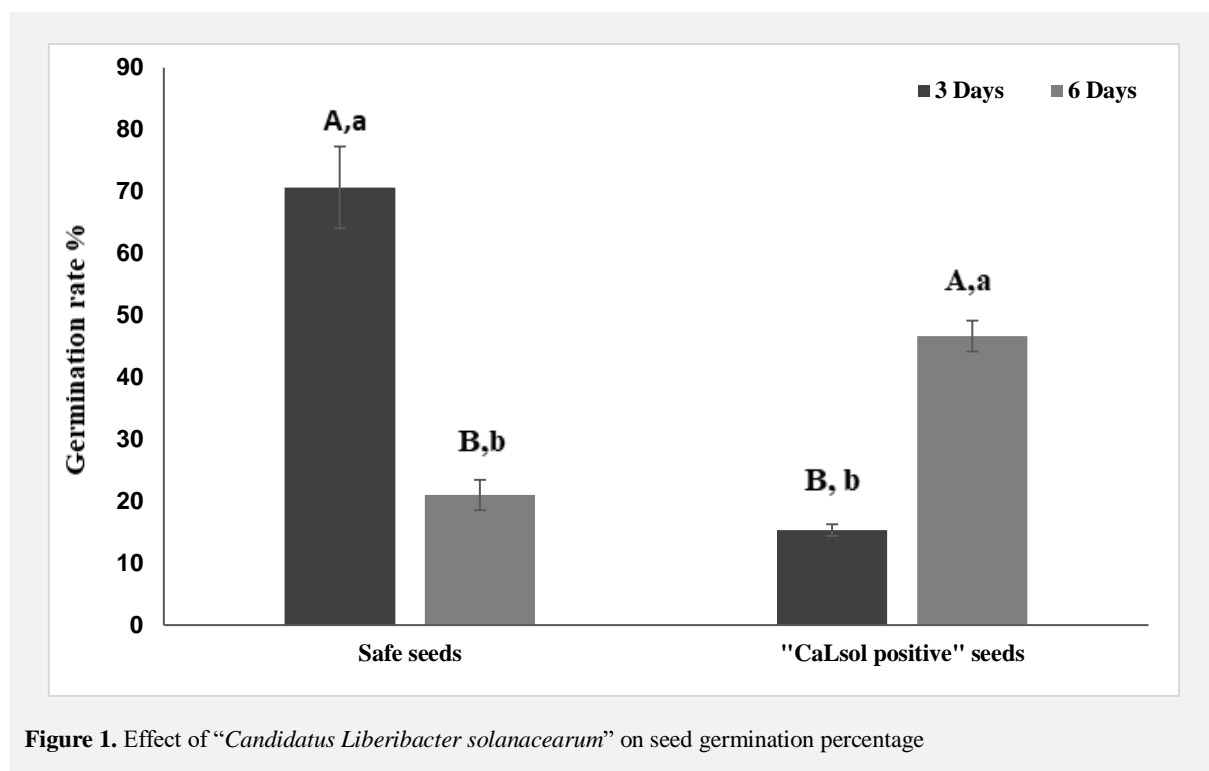
$$\text{GI} = \frac{\text{N0 of germinated seeds}}{\text{days of first count}} + \dots + \frac{\text{N0 of germinated seeds}}{\text{days of final count}}$$

### 2.4. Data analyses

Statistical analyses were performed using one-way analysis of variance ANOVA, and the significant difference between means was determined by Duncan’s multiple range test using SPSS 20 software. Significance was defined at  $P < 0.05$ .

## 3. Results and discussion

In present experiment the negative effect of the fastidious bacterium “*Ca. Liberibacter solanacearum*” on carrot seed germination rate was very clear. In fact, the germination rate recorded for the safe carrot seeds was significantly higher compared to the recorded one for contaminated seeds (Figure 1).



**Figure 1.** Effect of "*Candidatus Liberibacter solanacearum*" on seed germination percentage

Because of the presence of CaLso, this rate decrease significantly from 89.33% to 59.33% ( $P < 0.05$ ) for the contaminated carrot seeds (Table 1). Moreover, it can be postulated that this bacterium decreased the total number of germinating seeds and consequently the germination capacity. On the 3<sup>rd</sup> day after seeding we recorded a higher germination rate for the safe seeds (70.66%) while on the 6<sup>th</sup> day the highest germination rate was recorded for the CaLsol positive carrot seeds (40.66%). According to this result we can assume that the germination process was slowed down by the presence of bacterium which could explain the higher rate recorded on the 6<sup>th</sup> day for the contaminated carrot seeds.

Furthermore, the association of this bacterium to carrot seeds significantly decrease the germination index parameter ( $P < 0.005$ ). The mean germination time (MGT) was also significantly affected, by the association of CaLso 1 ( $P < 0.05$ ) (Table 1); however contaminated seeds required more time for germinating. Impairing of this parameters greatly explain that bacteria may damage or even destroy the essential seedling structures (Andersen et al. 1955; McGee 1979). The present result is in agreement with the study of (Islam 2005). In addition, it has been reported that bacteria reduce significantly the germination percentage of seeds (Jamadar et al. 2001; Gupta et al. 1989).

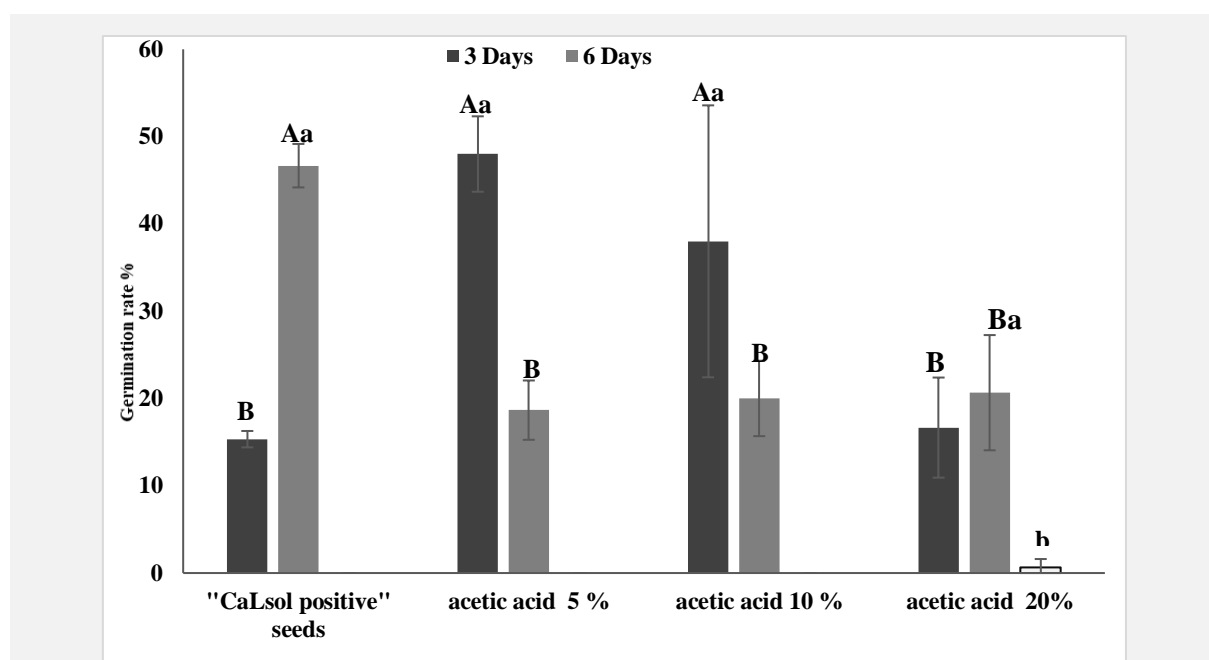
The observation of germination parameters of carrot seeds indicate that the safe seeds present the highest germination percentage and the highest germination index (Table 1) compared to the contaminated carrot seeds. This result unveiled the negative effect of this bacterium on the seed quality which consequently affects the germinations parameters specially the germination rate and the germination index. The CVG parameter was not significantly affected neither with the presence of CaLsol nor with the acetic acid treatment.

**Table 1.** Effect of treatment with acetic acid at different concentrations on germination parameters of carrot seeds-associated with "*Ca. Liberibacter solanacearum*"

Treatment	PG (%)	GI	CVG (d-1)	MGT (d)
Safe seeds	89.33 <sup>a</sup> (SE= 5.03)	19.66 <sup>a</sup> (SE=4.15)	0.387 <sup>a</sup> (SE=0.05)	2.64 <sup>a</sup> (SE=0.32)
"CaLsol positive" seeds	59.33 <sup>b</sup> (SE=5.03)	8.98 <sup>bc</sup> (SE=0.9)	0.29 <sup>a</sup> (SE=0.04)	3.47 <sup>bc</sup> (SE=0.51)
Acetic acid 5 %	66.66 <sup>b</sup> (SE=4.61)	11.78 <sup>b</sup> (SE=1.11)	0.325 <sup>a</sup> (SE=0.05)	3.09 <sup>ab</sup> (SE=0.06)
Acetic acid 10 %	59.33 <sup>b</sup> (SE=15.14)	9.39 <sup>bc</sup> (SE=3.49)	0.292 <sup>a</sup> (SE=0.044)	3.45 <sup>abc</sup> (SE=0.6)
Acetic acid 20%	38 <sup>c</sup> (SE=14)	5.52 <sup>c</sup> (SE=1.9)	0.34 <sup>a</sup> (SE=0.13)	3.91 <sup>c</sup> (SE=0.2)

The effect of different concentrations of acetic acid was studied on germination parameters of 'CaLsol' positive carrots seeds, since organic acids are known for their antibacterial and antifungal properties and especially that they were widely used in agriculture (De Muynck et al. 2004; Laitila et al. 2002; Lavermicocca et al. 2003; Pao et al. 2008; Sathe et al. 2007; Scholberg and Gaunce 1995; 1996, Tripathi and Dubey 2004). Furthermore, acetic acid is considered as a soft natural substance having the minimum toxicity risk for humans and animals (Borgen 2001) and it was usually used to reduce seed-associated micro-organisms.

Results showed that, compared to control, the germination rate of treated carrot seeds was higher on the third day. The untreated seeds lot exhibited a germination rate of 15.33%. When treated with the first concentration of acetic acid (5%), we record a value of 48% of normal seedling. At 10% acetic acid we observed an improvement in the number of germinated carrot seeds (38%) but, with a proportionally low number compared to that recorded for the first concentration. According to this result we could note that acetic acid has a simulative effect on germination ability. A negative effect of acetic acid on germination rate was observed on the 6<sup>th</sup> day after seeding, actually the use of acetic acid decreases the germination rate of the contaminated seeds. The observed results may greatly explain that the use of acetic acid at an adequate concentration has a significant effect in germinating seeds, particularly in increasing the number of germinating carrot seeds in early days such as on 3<sup>rd</sup> day after seeding.



**Figure 2.** The effect of different concentration of acetic acid treatments on germination percentage of carrot seed infected or not with "*Ca. Liberibacter solanacearum*"

When acetic acid was applied at a concentration of 20% results showed a significant decrease in the germination rate. This may explained by the fact that higher concentration of acetic acid could have

negative effect on seed germination ability. In addition the obtained results revealed that the higher concentration of acetic acid (20%) prolonged the germination of carrot seeds (Figure 2). Similar result was found by Szopińska (2013), when she studied the effect of organic acid on germination of Zinnia seeds.

Results of the seed treatments indicate that acetic acid has antimicrobial properties and it is able to reduce the life cells of associated bacteria which may explain the enhancement of the germination rate compared to control treatment. Indeed, Sholberg and Gaunce (1995) mentioned that low concentration of acetic acid vapour were extremely effective for the control of *Botrytis cinerea* conidia on apple fruit. Furthermore, (Borgen and Kristenson 2001) noted the positive effect of acetic acid for the control of common bunt (*Tilletia tritici*) in wheat. In addition, this treatment was also efficient for the control of *Alternaria dauci* in carrot (Lizot et al. 2002). The broad-spectrum antimicrobial activity of acetic acid has been also demonstrated by Lang et al (2000) as they found that acetic and lactic acid combined with hypochlorite may eliminate *Escherichia coli* O157:H7 from alfalfa seeds prior sprouting. In fact, Pao et al. (2008) achieved elimination of *Salmonella enterica* in alfalfa and mung bean sprouts after 4 and 16 hours immersing in 5% acetic acid. Further studies realized on cabbage seed-associated bacteria demonstrated that disinfection with lactic acid at 0.5% resulted in the reduction of the bacterial count by more than 99% at concentrations of 2.5 % (Van der Wolf et al. 2008). Besides, a high concentration ( $\geq 2\%$ ) of organic acids, (acetic acid, propionic acid, ascorbic acid and lactic acid) is required for a high antibacterial effect in the seed treatments (Van der Wolf et al. 2008).

#### 4. Conclusion

Following to these promising results further studies on carrot seeds treatment are needed with the aim of maintaining the benefit effects of acetic acid treatment, without affecting germination parameters. Then, optimization of acetic acid concentration for carrot seeds treatment against CaLso is therefore essential.

#### 5. References

- Andersen A M. (1955)** The effect of certain fungi and gibberellin on the germination of Merion Kentucky bluegrass seed. Proc. Assoc. Off. Analysts 45:145-153.
- AOSA (1983)** Association of Official Seed Analysis. Seed Vigor Testing Handbook. Contribution No. 32 to the handbook on Seed Testing.
- Ben Othmen S, Morán FE, Navarro I, Barbé S, Martínez C, Marco-Noales E, Chermiti B, López MM (2018)** 'Candidatus *Liberibacter solanacearum*' haplotypes D and E in carrot plants and seeds in Tunisia J P Pathol 100:197–207.
- Bertolini E, Teresani GR, Loiseau M, Tanaka FAO, Barbé S, Martínez MC, Gentit P, López MM, Cambra M (2014)** Transmission of 'Candidatus *Liberibacter solanacearum*' in carrot seeds. Plant Pathol 64:276–285.
- Bloukas JG, Paneras ED, Fournitzis GCA (1997)** Sodium lactate and protective culture effects on quality characteristics and shelf-life of low-fat frankfurters produced with olive oil. Meat Science 45: 223- 238.
- Borgen A (2004)** Strategies for regulation of seed borne diseases in organic farming. Seed Testing International 127 : 19-21.
- Borgen A, Kristensen L (2001)** Effect of seed treatment with acetic acid in control of seed borne diseases. In: Proceedings from BCPC Symposium No. 76: "Seed Treatment: Challenges & Opportunities" (Biddle, A.J., ed.) BCPC, Farnham, 135-140.
- De Muynck CD, Leroy AIJ, Maeseneire SD, Arnaut F, Soetaert W, Vandamme EJ (2004)** Potential of selected lactic acid bacteria to produce food compatible antifungal metabolites. Microbiol. Res. 159 : 339–346.
- El-Naimi M, Toubia-Rahme H, Mamluk OF (2000)** Organic seed-treatments as a substitute for chemical seed-treatment to control common bunt of wheat. Eur J P Pathol 106, 433-437
- Gamelin FX, Baquet G, Berthoin S, Thevenet D, Nourry C, Nottin S, Bosquet L (2009)** Effect of high intensity intermittent training on heart rate variability in prepubescent children. Eur J Appl Physiol 105:731-738. doi: 10.1007/s00421- 008-0955-8.
- Gupta k Sindu IR, Nazz S (1989)** Seed mycoflora of *Abelmoschus esculentus* (L.) Moench: A Survey and enumeration. Journal of Acta Botanica Indica 17: 200-206.
- Haapalainen M (2014)** Biology and epidemics of *Candidatus Liberibacter* species, psyllid-transmitted plant-pathogenic bacteria. Ann Appl Biol 165:172–198.



- Hansen A, Trumble J, Stouthamer R, Paine T (2008)** A new huanglongbing species “*Candidatus liberibacter psyllaurosus*,” found to infect tomato and potato, is vectored by the psyllid bactericera cockerelli (sulc). *Applied and environmental microbiology* 74: 5862-5865.
- Islam MR (2005)** Studies on the health and seedling diseases of some leafy vegetables, MS Thesis, Department of Plant Pathology, Bangladesh Agricultural University, Mymensingh pp. 67
- Jamadar MM, Ashok S, Shamrao J, Sajjan A, Jahangidar S (2001)** Studies on seed mycoflora and their effect on germination of colour graded okra [*Abelmoschus esculentus* (L.) Moench]. *J Crop Res* 22: 479-484
- Laitila A, Alakomi H-L, Raaska L, Mattila-Sandholm T, Haikara A (2002)** Antifungal activities of two *Lactobacillus plantarum* strains against *Fusarium* moulds in vitro and in malting barley. *J. Appl. Microbiol.* 93: 566–576.
- Lang MM, Ingham BH, Ingham SC (2000)** Efficacy of novel organic acid and hypochlorite treatments for eliminating *Escherichia coli* O157:H7 from alfalfa seeds prior to sprouting. *Int. J. Food Microbiol.* 58:73–82.
- Lavermicocca P, Valerio F, Visconti A (2003)** Antifungal activity of phenyllactic acid against molds isolated from bakery products. *Appl. Environ. Microbiol.* 69(1):634–640.
- Liefting LW, Weir BS, Pennycook SR, Clover GRG (2009)** ‘*Candidatus Liberibacter solanacearum*’, a *Liberibacter* associated with plants in the family Solanaceae. *Int J Syst Evol Microbiol* 59:2274–2276
- Lizot JF, Griboval B, Guenard M (2002)** Mise au point d’une technique de désinfection des semences applicable en agriculture biologique—*Alternaria dauci* sur semences de carottes. In: 2ème Conférence internationale sur les moyens alternatifs de lutte contre les organismes nuisibles aux végétaux. Lille, 4-7 March, 2002. p. 1-7.
- McGee DM (1979)** *Penicillium* contamination of grass seed germination tests. *J. Seed Technol.* 4:2, 18-23.
- Pao S, Kalantari A, Khalid MF (2008)** Eliminating *Salmonella enterica* in alfalfa and mung bean sprouts by organic acid and hot water immersions. *J. Food Process. Preserv.* 32: 335–342.
- Saidi B, Azmeh F, Mamluk OF, Sikora RA (2001)** Effect of seed treatment with organic acids on the control of common bunt (*Tilletia tritici* and *T. laevis*) in wheat. *Mededelingen Faculteit Landbouwkundige en Toegepaste Biologische Wetenschappen Universiteit Gent*, 66: 213-221.
- Sathe SJ, Nawani NN, Dhakephalkar PK, Kapadnis BP (2007)** Antifungal lactic acid bacteria with potential to prolong shelf-life of fresh vegetables. *J. Appl. Microbiol.* 103: 2622–2628.
- Sholberg PL, Gaunce AP (1996)** Fumigation of high moisture seed with acetic acid to control storage mold. *Can. J. Plant Sci.* 76: 551–555.
- Sholberg PL, Gaunce AP (1995)** Fumigation of fruit with acetic acid to prevent post harvest decay. *HortScience* 30: 1271–1275.
- Slifka MK, Whitton JL (2000)** Clinical implications of dysregulated cytokine production. *J Mol Med.* doi:10.1007/s001090000086
- Szopińska D (2013)** the effects of organic acids treatment on germination, vigour and health of zinnia (*Zinnia elegans* Jacq.) seeds *acta sci. pol., hortorum cultus* 12(5): 17-29.
- Tripathi P, Dubey NK (2004)** Exploitation of natural products as an alternative strategy to control postharvest fungal rotting of fruit and vegetables. *Postharv. Biol. Tec.* 32: 235–245.
- Van der Wolf JM, Birnbaum Y, van der Zouwen PS, Groot SPC (2008)** Disinfection of vegetable seed by treatment with essential oils, organic acids and plant extracts. *Seed Sci. Technol.* 36: 76–88.